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# **Biosystematics of entomopathogenic nematodes: current status and future directions**

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and

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# Systematics

**“... not only classifies and names diversity of the organic world, but seeks to understand the processes that produce and sustain that diversity.”**

**Quicke, 1983**

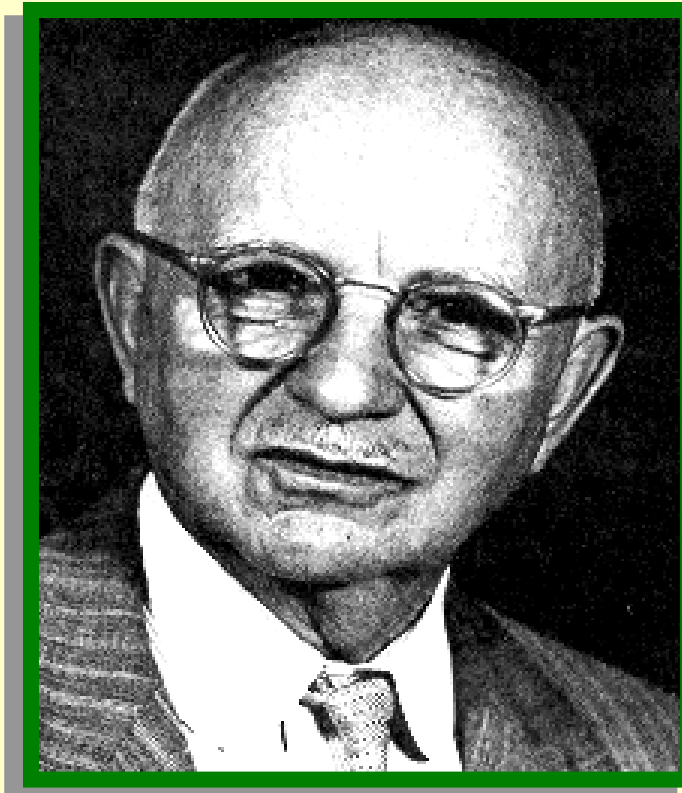


**Hmm...  
What kind of  
nematode  
is this?**



# Gotthold von Steiner

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1886-1961

# Family Steinernematidae Chitwood & Chitwood, 1937

## type genus: Steinernema Travassos, 1927

1. S. krausseii (Steiner, 1923) Travassos, 1927
2. S. glaseri (Steiner, 1929) Wouts, Mracek, Gerdin & Bedding, 1988
3. S. feltiae (Filipjev, 1934) Wouts, Mracek, Boemare, 1994
4. S. affine (Bovien 1937) Wouts, Mracek, Gerdin & Bedding, 1982
5. S. carpocapsae (Weiser, 1955) Wouts, Mracek, Gerdin & Bedding, 1982
6. S. arenarium (Artyukhovsky, 1967) Wouts, Mracek, Gerdin & Bedding, 1982
7. S. intermedium (Poinar, 1985) Mamiya, 1988
8. S. rarum (de Doucet, 1986) Mamiya, 1988
9. S. kushidai Mamiya, 1988
0. S. ritteri de Doucet & Doucet, 1990
1. S. scapterisci Nguyen & Smart, 1990
2. S. caudatum Xu, Wang & Li, 1991
3. S. longicaudum Shen & Wang, 1992
4. S. neocurtillae Nguyen & Smart, 1992
5. S. riobrave Cabanillas, Poinar & Raulston, 1994
6. S. cubanum Mracek, Hernandez & Boemare, 1994
7. S. puertoricense Roman & Figueroa, 1994
18. S. bicornutum Tallosi, Peters & Ehlers, 1995
19. S. monticolum Stock, Choo & Kaya, 1996
20. S. oregonense Liu & Berry, 1996
21. S. abbasi Elawad, Ahmad & Reid 1997
22. S. ceratophorum Heng, Reid & Hunt, 1997
23. S. kariii Waturu, Hunt & Reid, 1997
24. S. siamkayai Stock, Sommsook & Reid, 1998
25. S. tami Van Luc, Nguyen, Reid, Spiridonov, 2000
26. S. thermophilum Ganguly & Singh, 2000
27. S. sangi Ke, Nguyen & Moens, 2001
28. S. loci Ke, Nguyen & Moens, 2001
29. S. thanhi Ke, Nguyen & Moens, 2001
30. S. pakistanense Shahina, Anis, Reid, Rowe and Maqbool, 2001

## Neosteinerema Nguyen & Smart, 1994

1. N. longicurvicauda Nguyen & Smart, 1994

# Family Heterorhabditidae Poinar, 1976

Type Genus: Heterorhabditis Poinar, 1976

1. Heterorhabditis bacteriophora Poinar, 1976  
syn. Chromonema heliothidis Khan, Brooks & Hirschmann, 1976  
Heterorhabditis heliothidis (Khan, Brooks & Hirschmann, 1976)  
Poinar, Hess & Thomas, 1977
2. H. megidis Poinar, Jackson & Klein, 1988
3. H. zealandica Poinar, 1990
4. H. indica Poinar, Karunakar & David, 1992  
syn. H. hawaiiensis Gardner, Stock & Kaya, 1994
5. H. argentinensis Stock, 1993
6. H. brevicaudis Liu, 1994
7. H. marelatus Liu & Berry, 1996  
syn. H. hepialius Stock, Strong & Gardner, 1996
8. H. poinari Kakulia & Mikaia, 1997
9. H. downesi Stock, Griffin and Burnell, 2002

# Description Criteria in Currently Described EPN Species

☞ Phenetic species concept → Morphology

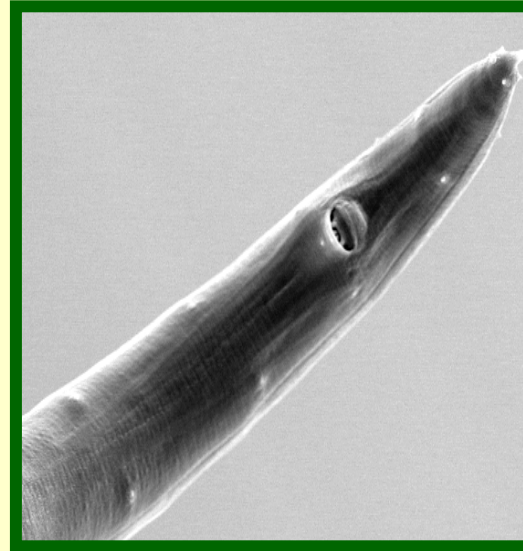
☞ Biological species concept → Cross-breeding tests

# Key Diagnostic Features

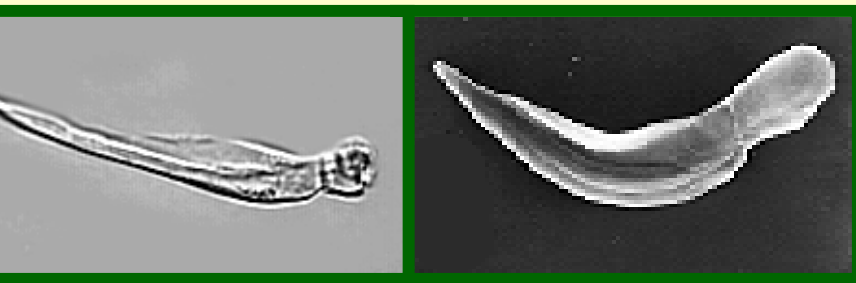
## 1st. Generation Male



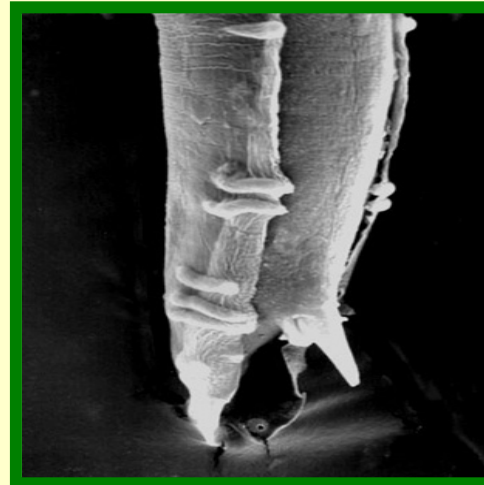
Gubernaculum



Tail,  
Genital papillae

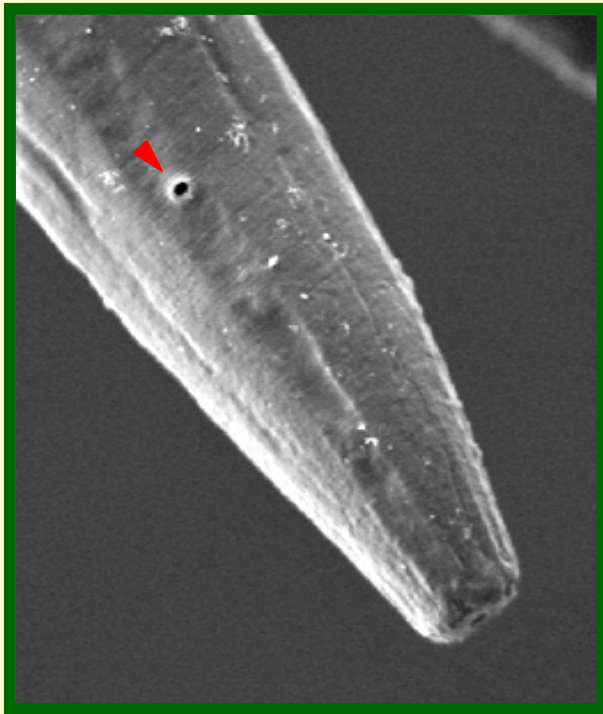


Spicules

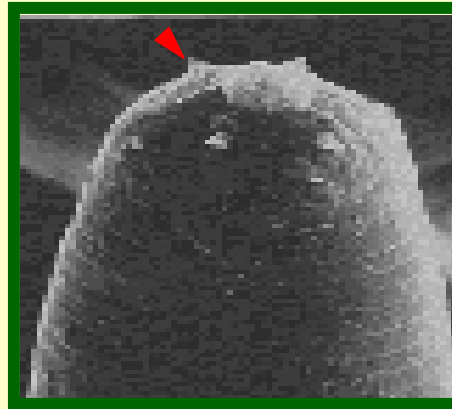


# Key Diagnostic Features

rd Stage Infective Juvenile:



Excretory pore location



Cephalic oddities



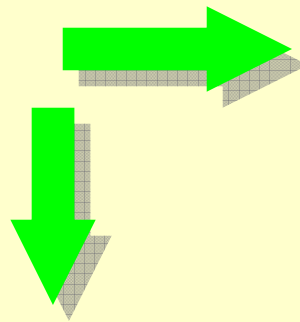
Lateral field pattern

# Limitations of currently used methods

Lack of morphological variation at the specific level.

Most morphological diagnostic traits have no evolutionary value because they represent either plesiomorphic states or are highly homoplasious characters.

Cross- Hybridization  
Methods




- ✓ Time consuming
- ✓ Too laborious

**A self-fertile species of *Steinernema* from Indonesia: further evidence of convergent evolution amongst entomopathogenic nematodes?**

C. T. Griffin, K. M. O'Callaghan and I. Dix. *Parasitology* (2001) 122, 181-186

# Molecular Approaches

- 
- **Protein electrophoresis:** Isozyme patterns } 1960's  
Total protein patterns } 1980's
  - **Immunological techniques:** Gel diffusion } 1980's,  
Immuno electrophoresis } 1990's
  - **RFLPs** } 1980's - present
  - **RAPDs** } 1990's - present
  - **Nucleotide sequence analysis** } late 1980's - present

# Assessment of Molecular Approaches for EPN Diagnostics



**Protein electrophoresis**

- Good at species level (sibling species)
- Little use above the genus level, or below the level of subspecies (unless levels of divergence are present)

**Immunological techniques**

- Useful at the species level



# Assessment of Molecular Approaches for EPN Diagnostics (cont.)

## **RAPDs**

- Useful at the subspecies level (populations, strains)

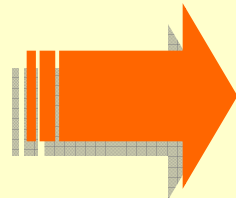
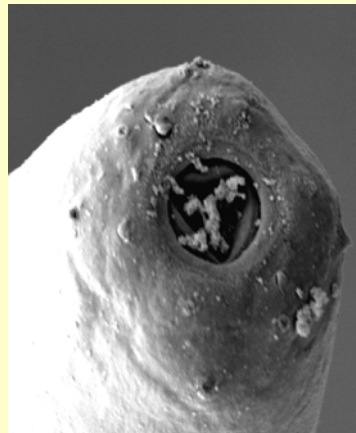
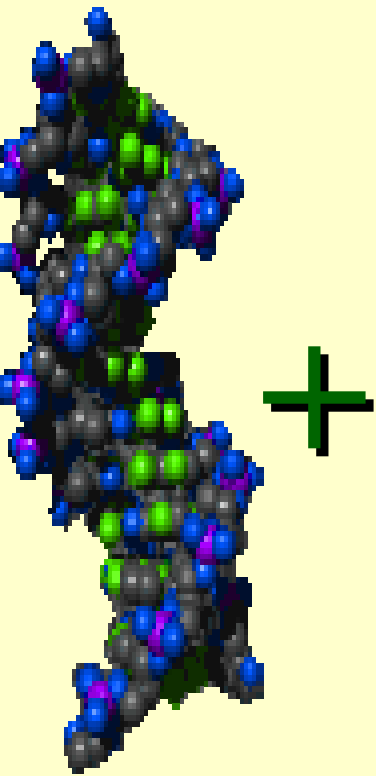
## **RFLPs**

- Useful at species levels and infra-specific categories

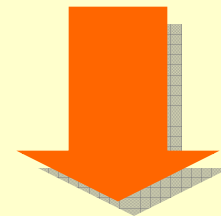
## **Nucleotide sequence analysis**

- Depending on the gene and gene-region targeted, this method can be valuable for discriminating taxa above, below or at the species level

# combined morphological-molecular approach.



**A more comprehensive  
view  
of EPN evolution**



**A better classification**

**Phylogeny of *Steinernema* Travassos, 1927 (Cephalobina: Steinernematidae) inferred from ribosomal DNA sequences and morphological characters. Stock, Campbell and Nadler, 2001. J. Parasitol. 87: 877-889.**

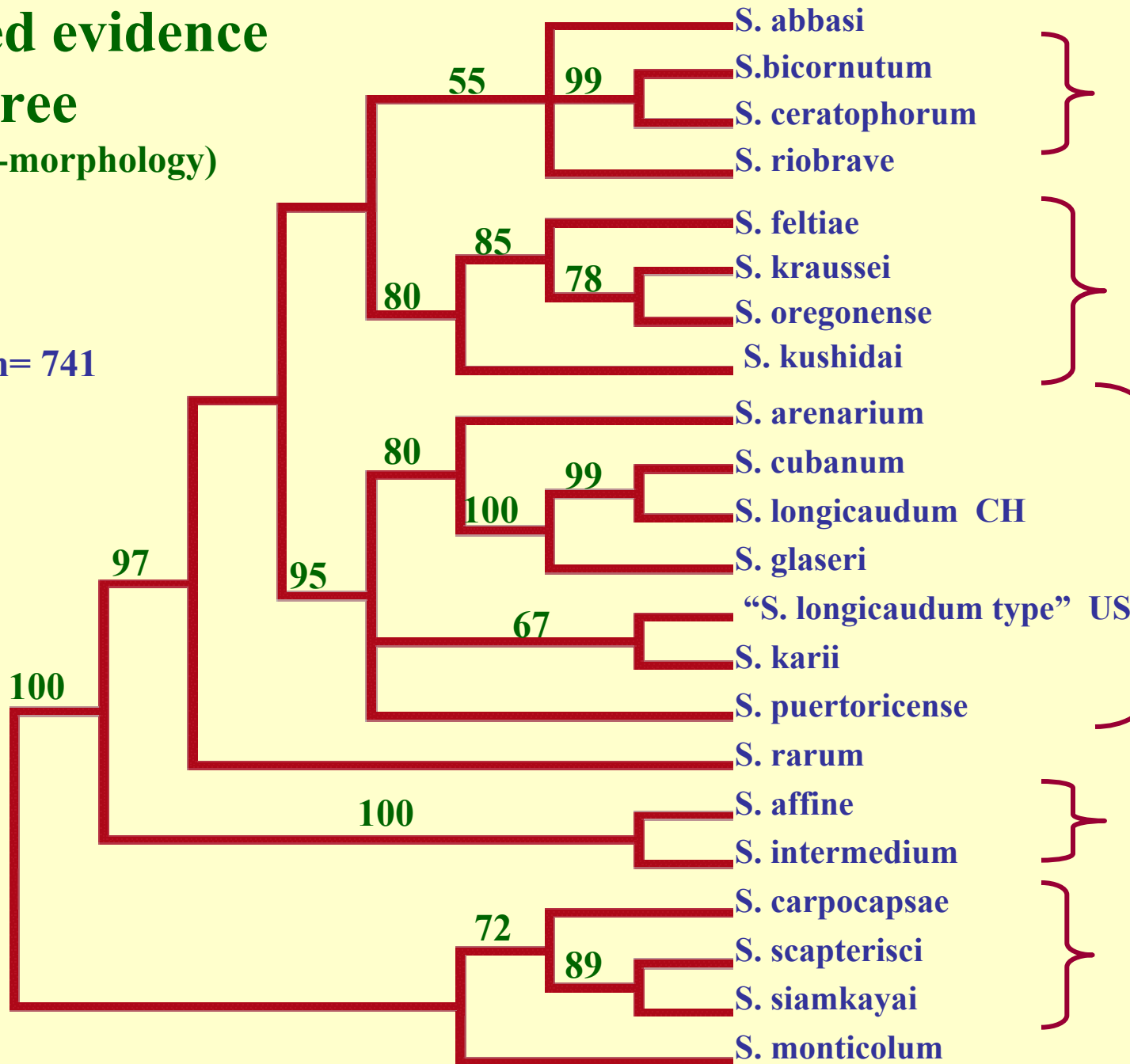
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- **Evolutionary relationships among *Steinernema* spp.**
- **Patterns of character evolution.**
- **Apply the “autapomorphy-base concept for delimitation of *Steinernema* species.**

# Combined evidence tree

(molecules-morphology)

Tree length= 741  
CI= 0.60



IJs with horn-like structure

Medium size IJ  
Average:  
650-900  $\mu$ m

Large IJ  
Average  
More than 900  $\mu$ m

Similar adult and IJ morphology

Small IJs  
Average:  
Less than 600  $\mu$ m

# Most of the morphological characters studied were...

**Homoplasious**

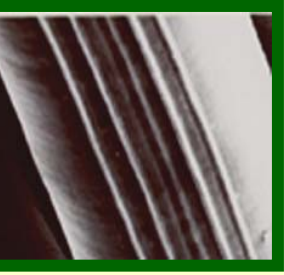


**non-homologous,  
variable results  
according to different  
tree-topologies**

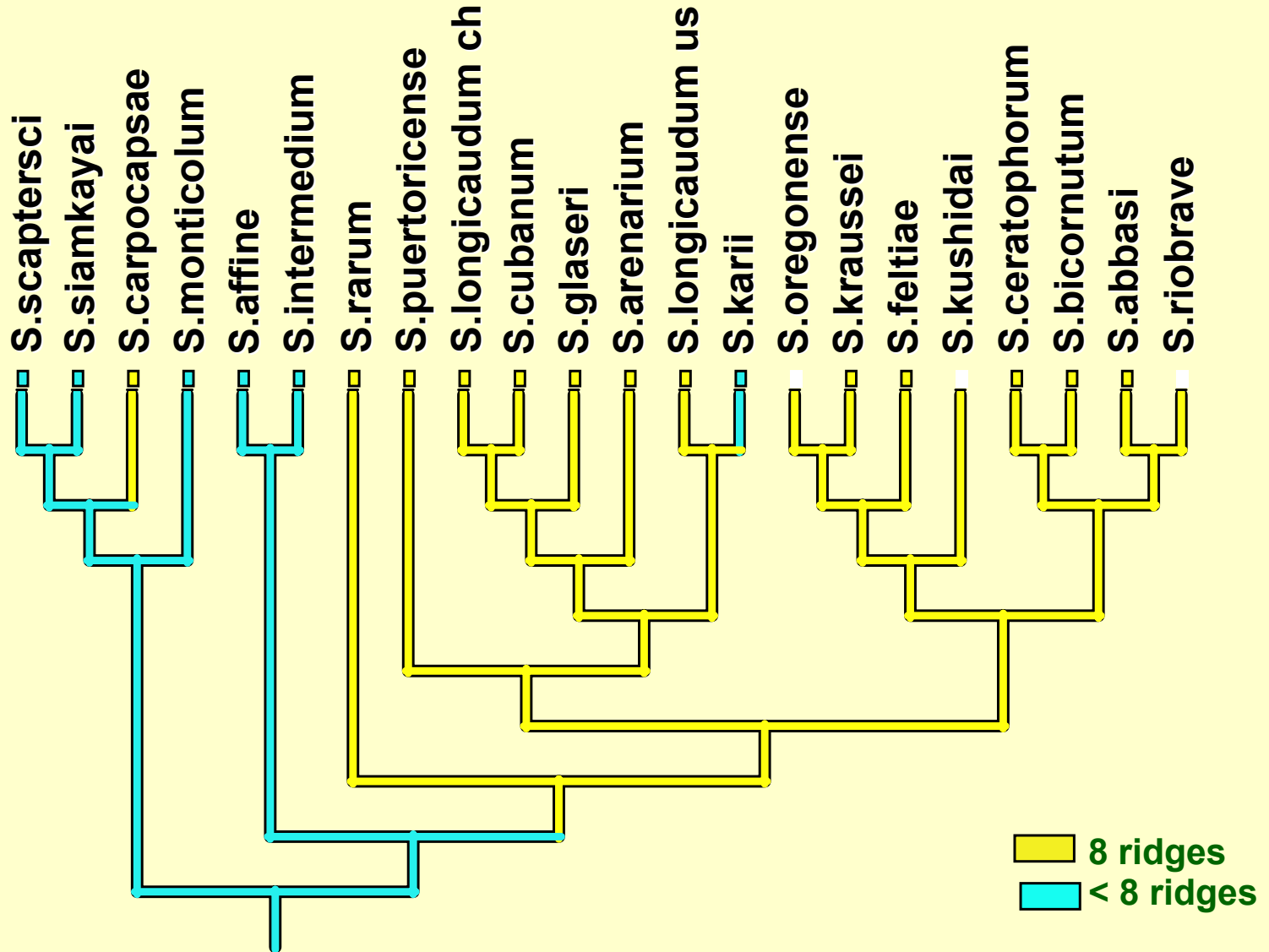
**Plesiomorphic**

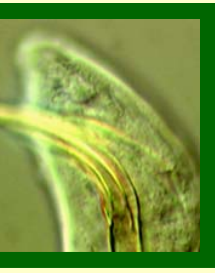


**ancestral states**

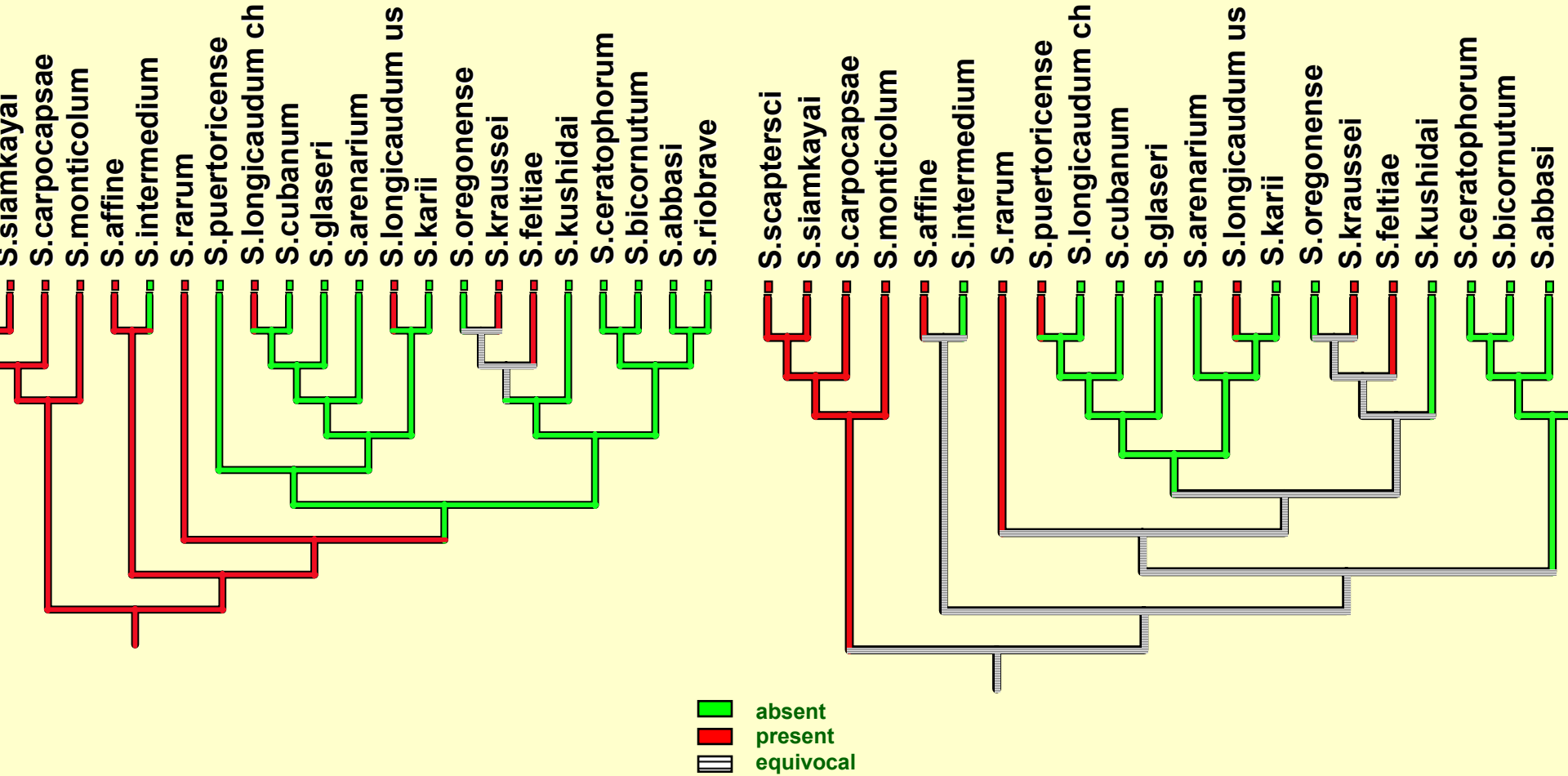


# IJ lateral field pattern



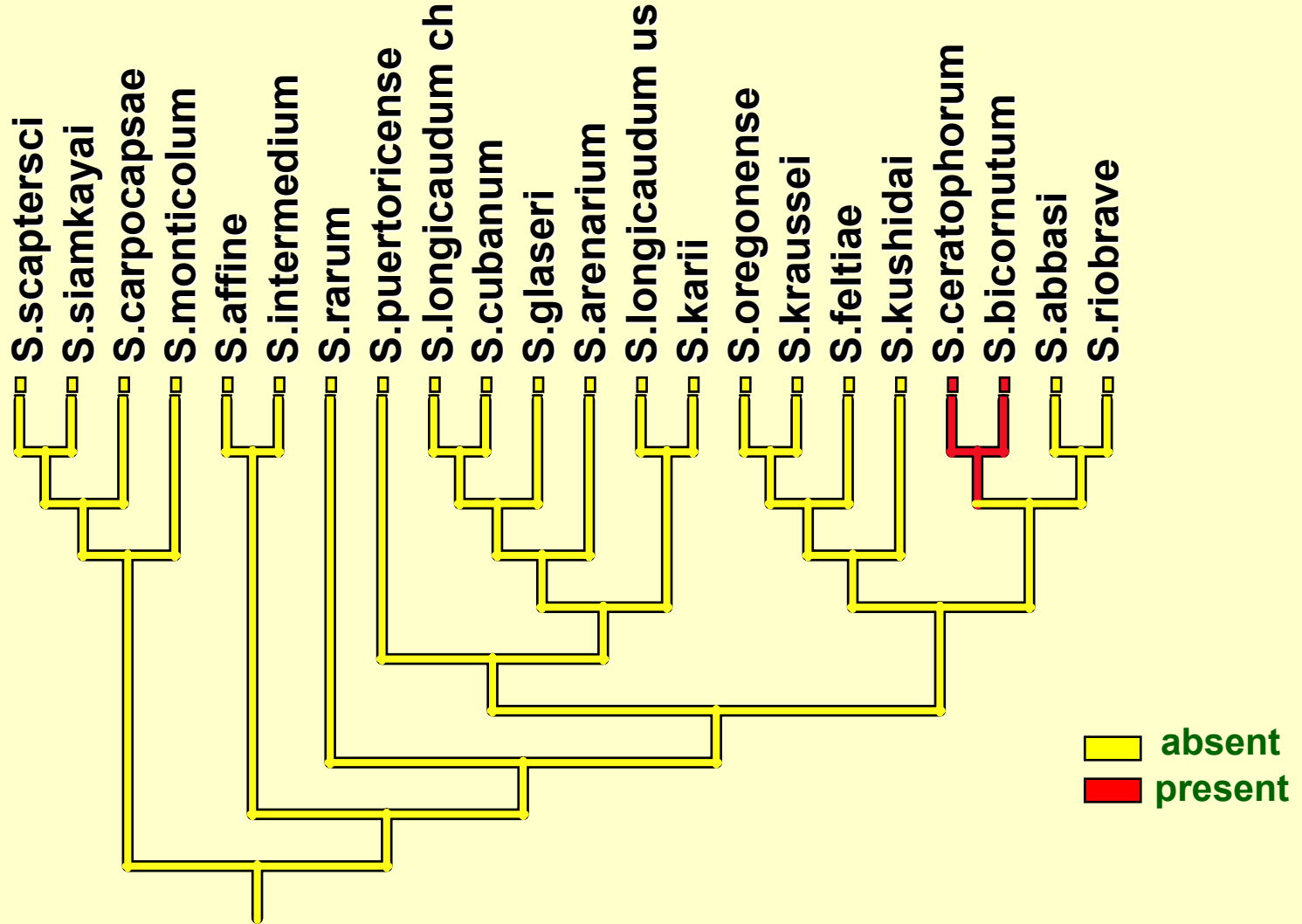


# Male:tail mucro





# IJ: Horn-like structures

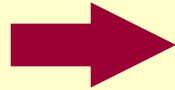


# Species Delimitation



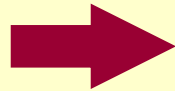
## “Automomorphy Concept”

➤ 4 combined evidence trees  
(morphology + LSU)



All taxa delimited by 3 or more autapomorphies

➤ 2 LSU trees  
(full alignment dataset)



All taxa delimited by 2 or more autapomorphies  
except *S. carpocapsae*

# Summarizing...

**This study provides a unifying framework where molecular and morphological tools have demonstrated to be complementary tools in EPN systematics.**

**This combined morphological-molecular approach has shown to be valuable not only for understanding evolutionary relationships among species in the genus *Steinernema*, but also for interpreting patterns of character evolution of morphological traits and in assessing diagnosable units in this genus.**

Introductory slide to Alex's presentation

# Single site sampling strategy

- 100 soil samples taken from a 100m<sup>2</sup> grid.
- Each sample baited with 3 *Galleria* larvae.
- Progeny from each larvae analyzed separately.
- Soil samples baited at several time points.
- Should give an indication of genetic diversity over space and time.

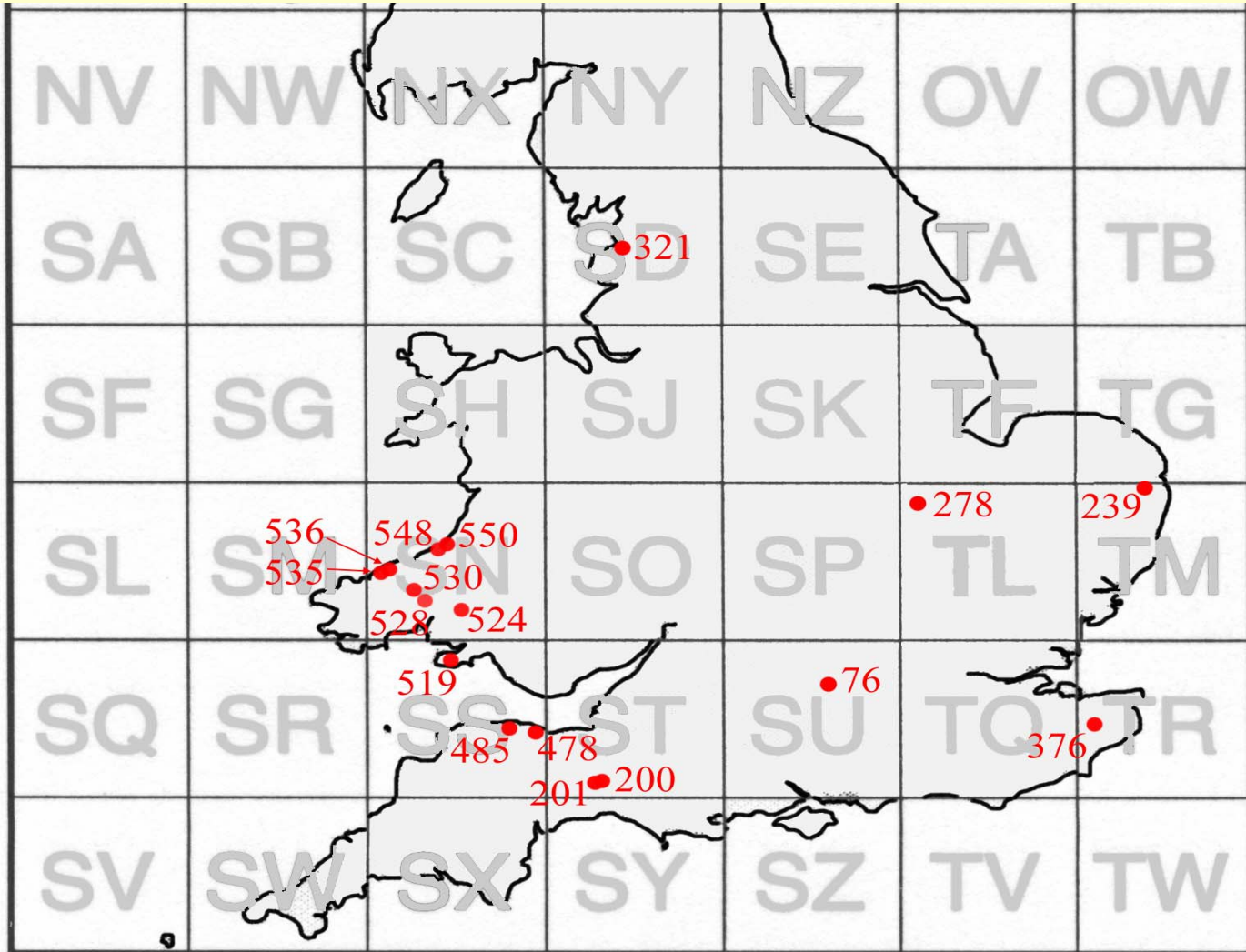
D'OH



# ***S. feltiae* sites**

<b>Site number</b>	<b>Grid reference</b>	<b>Habitat type</b>
76	SU 568 729	Hedgerow
200	ST 280 108	Field - Pasture
201	ST 277 088	Field - Pasture
239	TM 396 995	Field - Cultivated
278	TL 096 874	Wood - Deciduous
321	SD 414 453	Field - Cultivated
376	TR 114 484	Field - Cultivated
478	SS 936 458	Verge
485	SS 802 483	Moorland
519	SS 412 812	Field - Pasture
524	SN 387 213	Field - Pasture
528	SN 288 277	Field - Pasture
530	SN 221 343	Field - Pasture
535	SN 066 406	Verge
536	SN 072 424	Field - Pasture
548	SN 345 555	Verge
550	SN 366 572	Field - Pasture

# UK *S. feltiae* site locations



# Three fingerprinting techniques used

AFLP

RAPD

ISSR

# ISSR technique

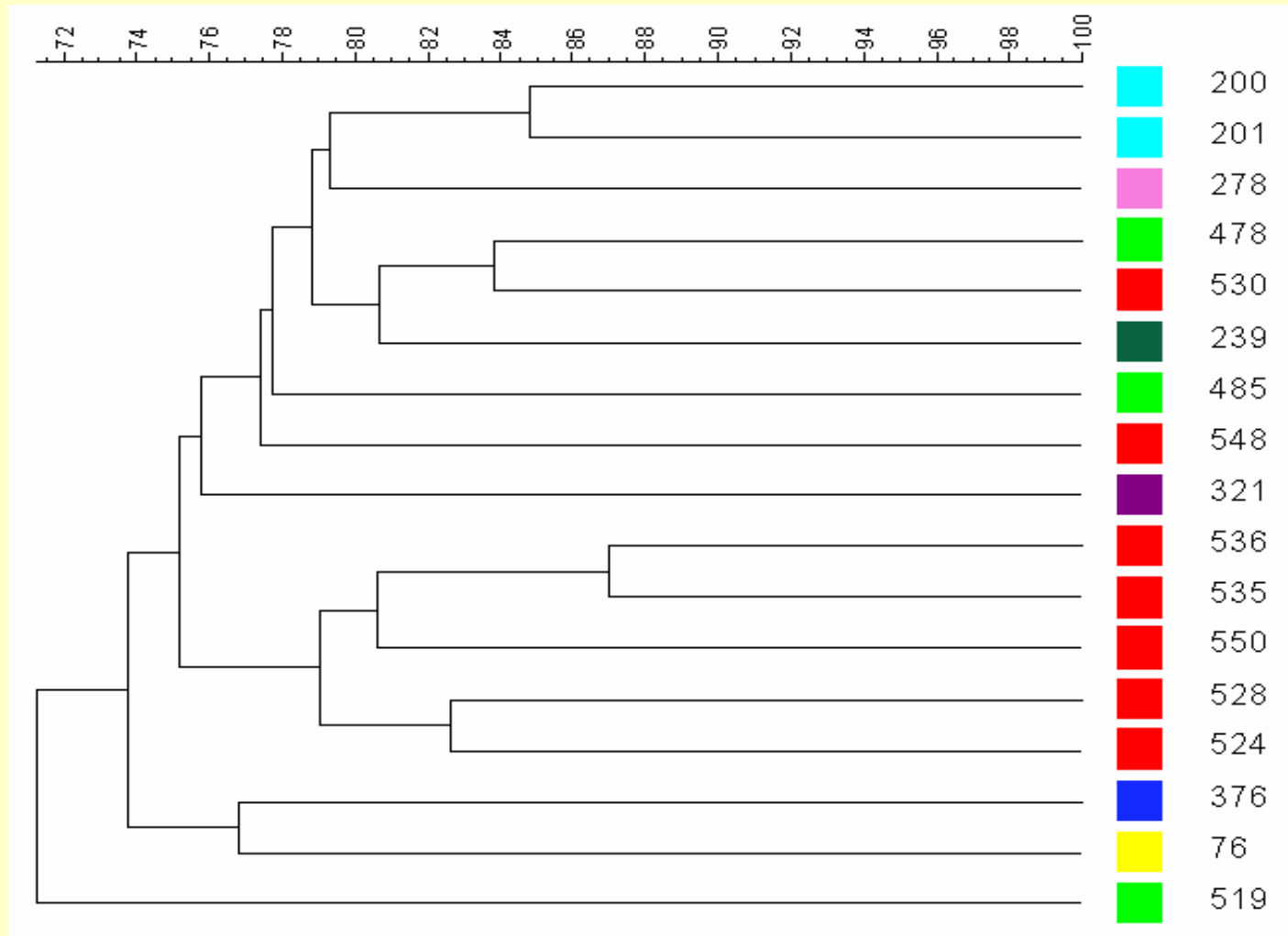
Interspersed Simple Sequence Repeats

- Primers have a core microsatellite sequence with a degenerate 5' end

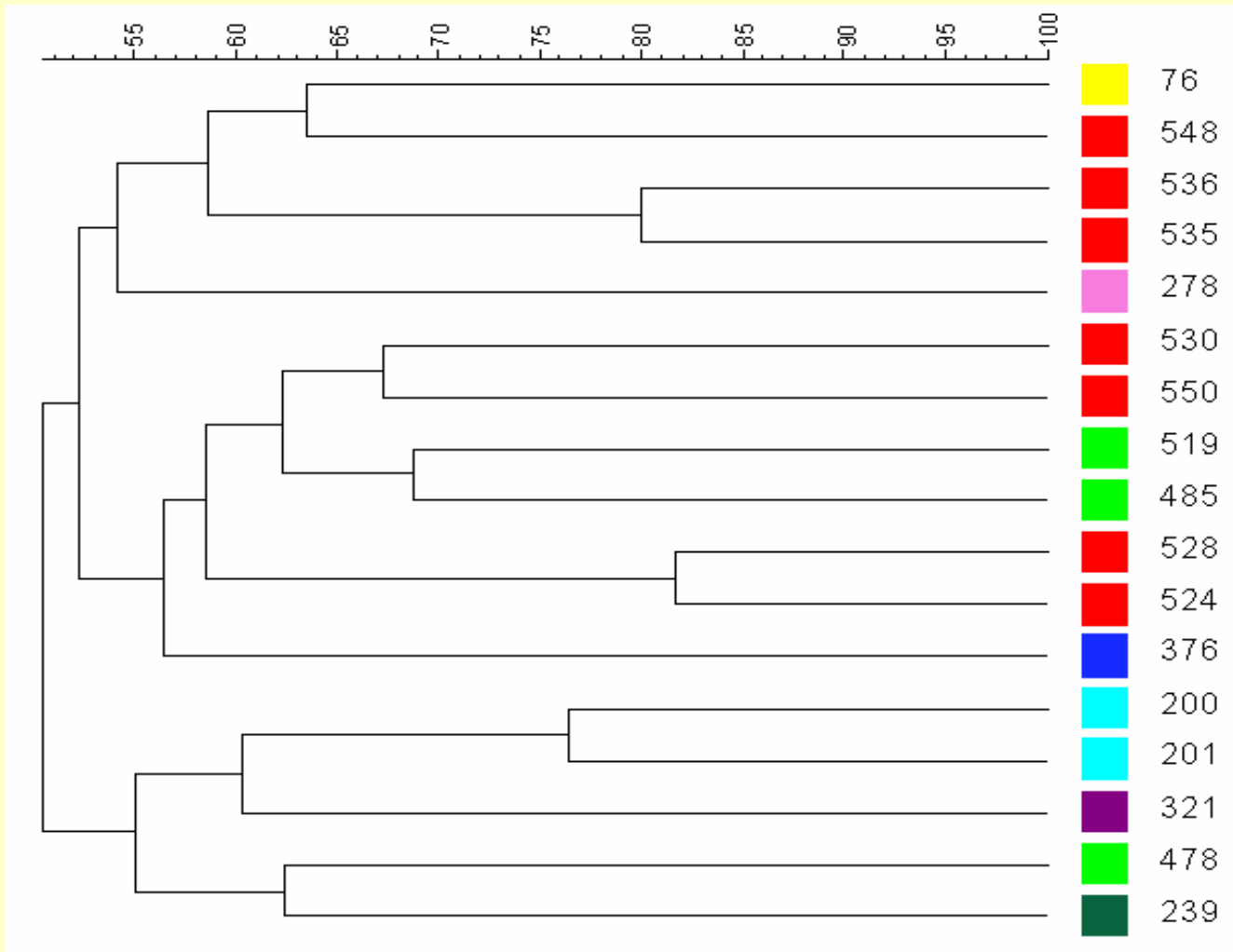
BDB (ACA)<sub>5</sub> or DD (CCA)<sub>5</sub>

- Amplify regions of genomic DNA flanked by microsatellite like sequences.
- Random distribution of microsatellite sequences in the genome yield polymorphic bands.

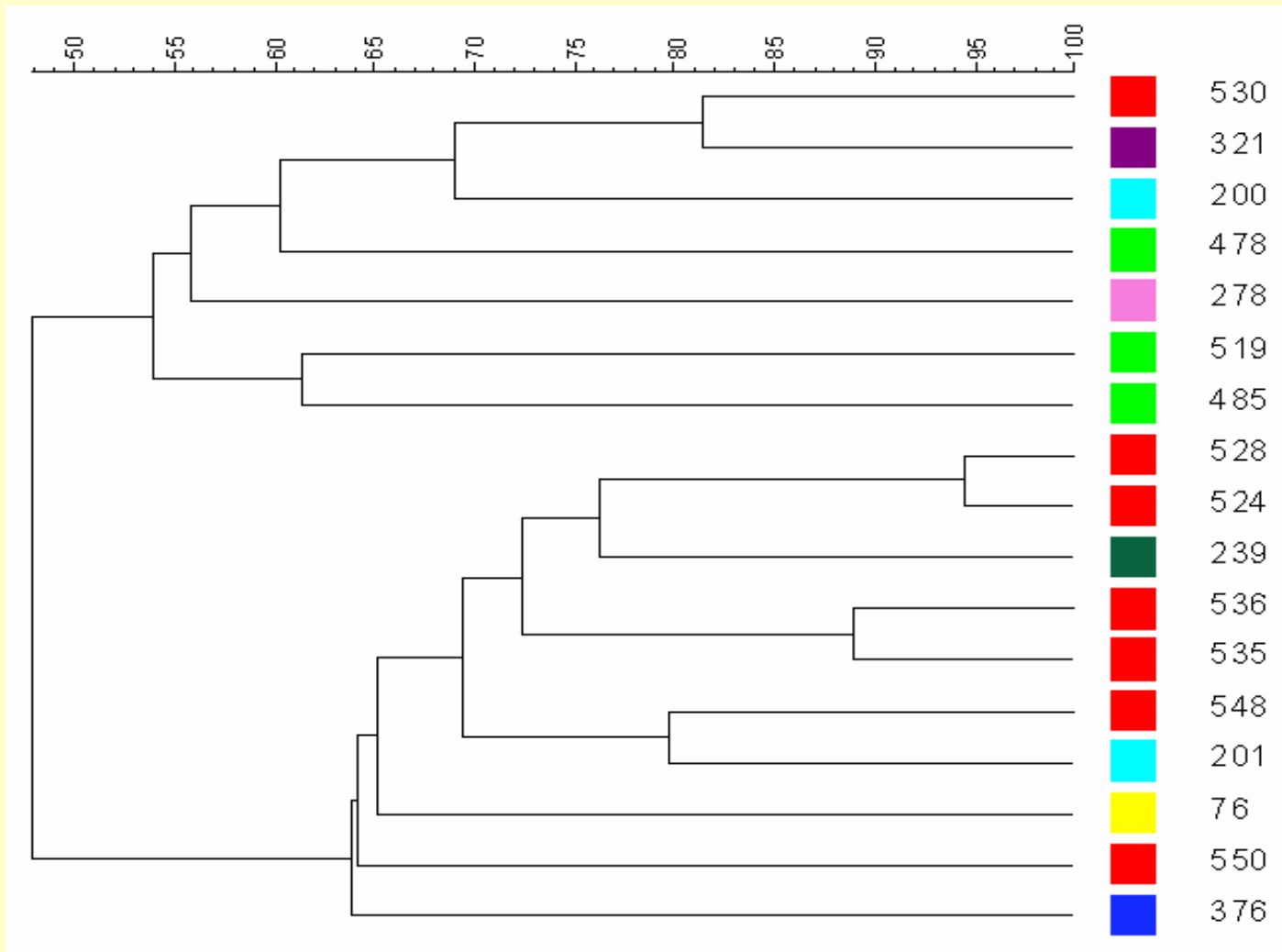
# *S. feltiae* (AFLP data)



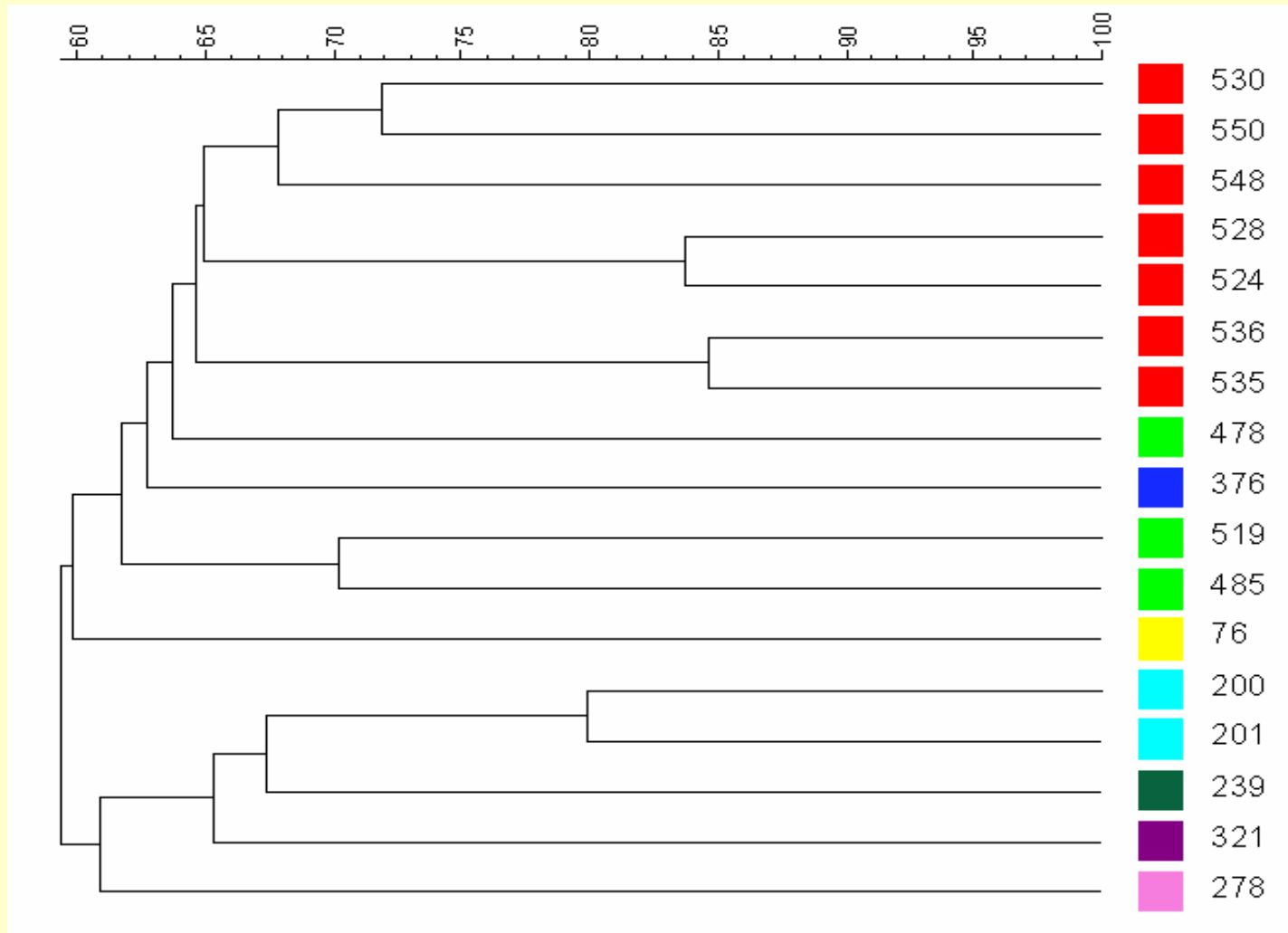
# *S. feltiae* (RAPD data)



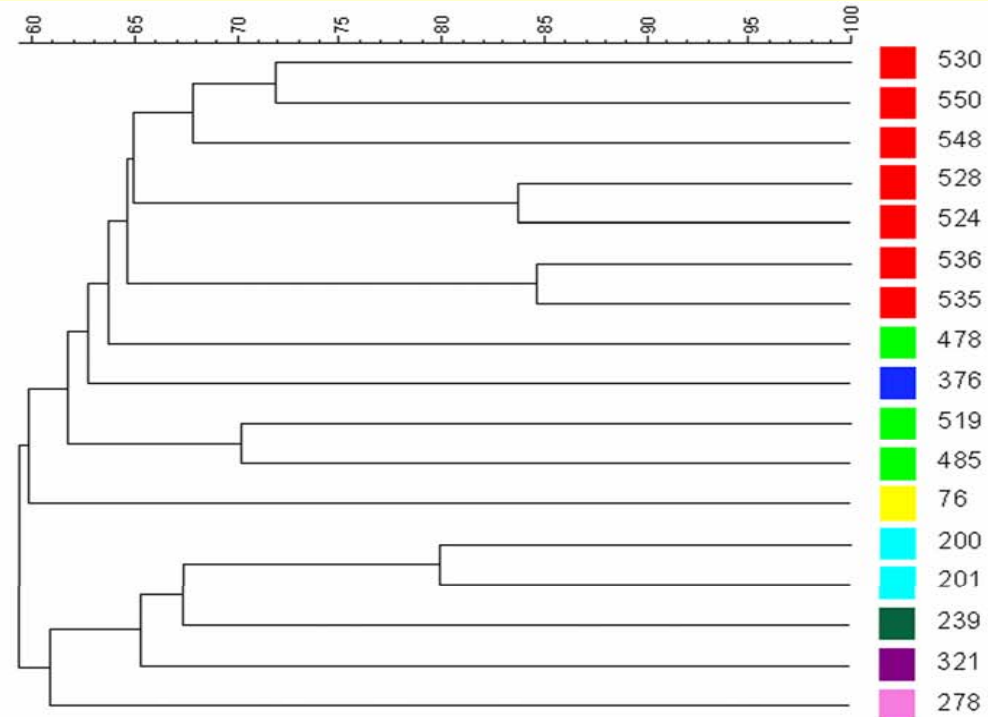
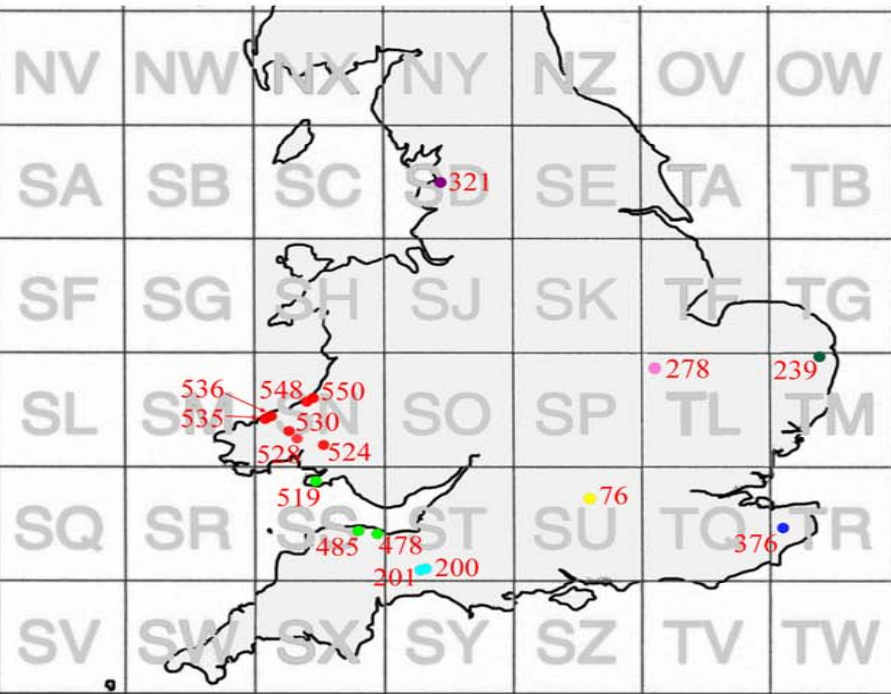
# *S. feltiae* (ISSR data)



# *S. feltiae* (combined data)



# *S. feltiae* (combined data)



# Conclusions - UK *S. feltiae* samples

- Three different fingerprinting approaches gave different results.
- No good correlation between geographical location and diversity for individual techniques.
- Correlation between geographical location and genetic diversity. (when data for all three methods combined).